

Diversity of the HrpF/NolX/PopF protein in plant pathogenic bacteria

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Introduction The interaction between the Gram-negative plant pathogenic bacteria and its host plants is controlled by *hrp* genes (*hypersensitive reaction and pathogenicity*), which encode a type III protein secretion system (T3SS). T3SS-secreted proteins include *hrp*-related translocators and effectors involved in plant defense suppression. The HrpF/NolX/ HrpK/ PopF1/ PopF2 family of *hrp* proteins (PF05819) includes *Rhizobium* NolX, *Ralstonia* Pop1/2, and *Xanthomonas* HrpF proteins [1]. Despite large taxonomic distance between genera *Rhizobium* (alpha-proteobacteria), *Ralstonia* (beta-proteobacteria), and *Xanthomonas* (Gamma-proteobacteria), HrpF, NolX and PopF1/ PopF2 have remarkably high similarity [3,4]. HrpF is a translocator of effector proteins into the host cell, dispensable for protein secretion but required for effector AvrBs3 recognition in planta [2]. NolX, a legumes cultivar-specific protein, is expressed in planta mostly during the early stages of nodule development [3], secreted by a T3SS [4]. Expression of *nolX* is induced as much as 30 fold by flavonoid signal molecules, even though these genes lack *nodbox* promoters [5]. It was speculated that HrpF protein may play a role in host-specialization of the bacteria [6], and *hrpF* gene -specific primers were successfully used for identification of *X. campestris* [7].

We evaluated protein diversity across different HrpF/NolX/PopF1/PopF2 group members available in the GeneBank, and sequenced nearly complete *hrpF* gene fragment for 65 strains of xanthomonads obtained from brassicas with symptoms of black rot or leaf spots across Russia and some related bacteria isolated from crops of other families at the same regions. 91 proteins of HrpF/NolX/HrpK/PopF1/PopF2 family were found in the GeneBank by BLAST search against HrpF protein of *Xanthomonas campestris* pv. *campestris* str. ATCC 3391372. 65 phenotypically and genetically characterized strains of *Xanthomonas* from brassicas collected in Russia, and

representative strains of related bacteria from USA, Japan, Germany, and other countries were obtained from collection of Russian Research Institute of Phytopathology. Majority of the strains belonged to *X. campestris* pv. *campestris*, pv. *raphani*, pv. *armoraciae*, and included some strains of *X. arboricola*, *X. vesicatoria*, and *X. hortorum*. Nearly complete DNA sequence of *hrpF* gene (85%) was obtained with two pairs of sequencing primers by direct PCR amplification [8].

Results The HrpF proteins obtained for *Xanthomonas* spp., NolX, and PopF1/ PopF2 had a length ranged from 806 to 921aa. The protein is hydrophilic, but it has two hydrophobic domains at the C-terminal end. The protein sequence does not contain signal peptides. Molecular weight is ranged from 87 to 98.9 kDa. A detailed analysis of the obtained amino acid sequence revealed two repetitive regions in HrpF/NolX/Pop1 proteins. One of these is a conserved domain with 118aa length. In most proteins of the HrpF/Pop1 group this conserved domain is present in two copies, but in the NolX proteins it is single-copy, and in the HrpF protein from *X. campestris* the 188aa domain is represented by three copies and demonstrates a possible origin from recombination with the homologous protein from *X. fuscans*.

The second repetitive region is a 4aa degenerate repeat (consensus sequence AKGA). The first 4 repeats of amino acid sequence **AKGA** are coded by conservative DNA sequence 5'-CTC GCA GCC AAG GGA GCA GCC AAG GGG GTC GCC GAG GGC GCA GCC AAG GGA GCG ACC CAG GGT-3' present in many sequenced bacterial genomes. Some regulatory proteins can bind these sequences. We also assume that these repeats are responsible for the rapid evolution of this tandem repetitive region of the HrpF protein and may determine the range of host-plants.

In the HrpF protein of *X. campestris* it starts at 810aa position, and present from 8 to 28 times in tandem repeats in the homologous protein sequences from other bacteria of the genus *Xanthomonas*. We have noted the similarity of the four amino acid repeat with the EKGI/AKGI repeat of transposase, which is probably responsible for the DNA binding. The analysis of the obtained *hrpF* gene nucleotide sequences from xanthomonads isolated from brassicas mainly reflected the host plant range of the isolated bacteria then their taxonomic position obtained by MLST analysis (data not shown).

Conclusions HrpF/NolX/Pop1 proteins are probable determinants of host range of the plant pathogenic and symbiotic bacteria with nucleotide binding activity and specific mechanism of their rapid evolution.

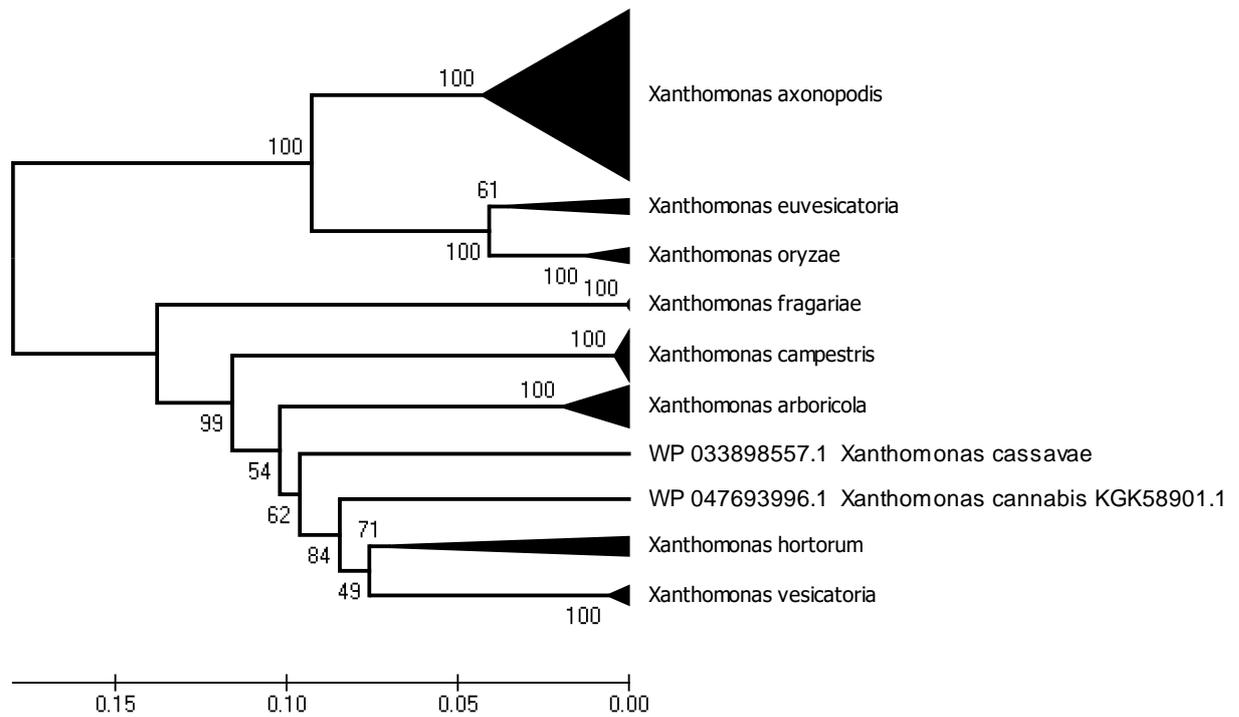
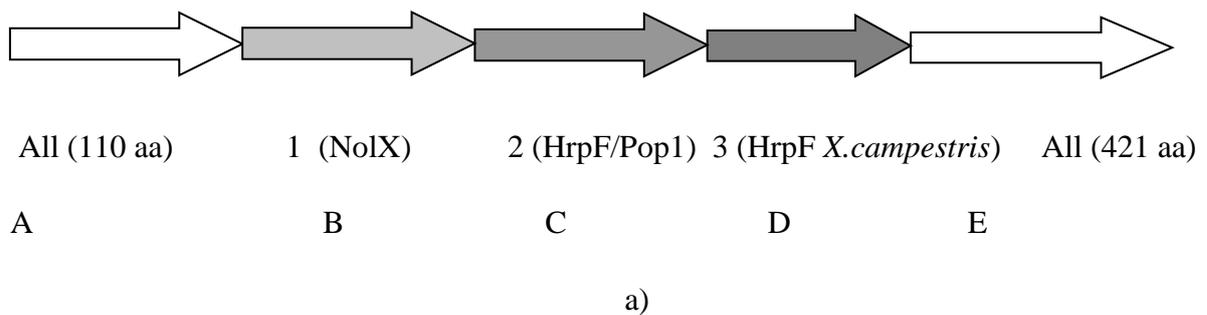


Figure 1. Evolutionary relationships of 91 proteins of HrpX/NolX/Pop1 group from *Xanthomonas* spp.. The evolutionary history was inferred using the Neighbor-Joining method. The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (1000 replicates) are shown next to the branches. Phylogenetic analyses were conducted in MEGA7.



>Consensus repeated sequence of HrpF protein:

F*PK*IKG*PPA**GS*VTW**GTLT*SEL*IVSTLN*HKD*****LD*KINDPSTPPD
 LK*AL*GL**DPRLFFAIGSQGDGKCGGK*****L*DF***H*Q*****K*

b)

Figure 2. Conservative and variable repeated domens (B,C and D) of HrpF/NolX/Pop1 protein (a), and consensus repeated sequence of HrpF protein (b).

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